SEVERE DAMAGE CAUSED BY THE ROOT-LESION NEMATODE, *PRATYLENCHUS THORNEI*, IN AEROBIC RICE IN INDIA

Pankaj¹, A.K. Ganguly¹ and R.N. Pandey²

¹Division of Nematology, ²Division of Soil Science and Agricultural Chemistry Indian Agricultural Research Institute, New Delhi – 110 012, India

Received: 19 March 2012; Accepted: 21 May 2012.

Summary. Severe damage to aerobic rice caused by the root lesion nematode, *Pratylenchus thornei*, was observed in India. Infected plants showed retarded growth, chlorotic leaves and roots with too many branches and necrotic lesions throughout the cortex, compared to apparently healthy plants of a nearby rice crop. The nematode populations were 406 specimens per 200 cm³ soil and 32 specimens per 0.5 g roots. This is the first report of *P. thornei* damaging aerobic rice.

Keywords: Oryza sativa, upland rice, yield loss.

In Asia, 75% of rice is produced in irrigated lowland fields with high irrigation requirements to sustain a layer of ponded water for most of the growing season (Bouman and Tuong, 2001; Maclean et al., 2002; Bridge et al., 2005). However, irrigation water is becoming increasingly scarce. Tuong and Bouman (2003) estimated that about 2 million ha of Asia's dry-season irrigated rice area will suffer water shortages by 2025. One of the adaptation strategies for water shortage areas is the system of aerobic rice (Bouman et al., 2005, 2006; Yang et al., 2005). Aerobic rice is a water-saving rice production system for water-short environments with favourable soils and adapted, potentially high-yielding varieties that are direct dry seeded. Soils remain aerobic (not saturated) but supplementary irrigation is applied as necessary. In light soils with high seepage and percolation rates, aerobic rice has an advantage over systems that include saturated or partially flooded soil conditions (Wang et al., 2002).

Lesion nematodes are widely distributed in the world and mainly inflict damage to direct-seeded rainfed rice. In India, *P. indicus* Das and *P. zeae* Graham have been recorded on rice in Andhra Pradesh, Assam, Gujarat, Kerala, Orissa, Madhya Pradesh, Rajasthan, Uttar Pradesh and West Bengal (Prasad *et al.*, 1987). Damage by *P. thornei* to chickpea (Di Vito *et al.*, 1992), mung bean and wheat (Nicol *et al.*, 1999; Nicol and Ortiz-Monasterio, 2004) have been reported. The nematode has already been reported to damage the rice-wheatlegume cropping sequence in India (Dwivedi and Upadhyay, 1998), but never upland rice. In rice, *P. thornei* was first reported by Oteifa (1962) from Egypt.

Lately, rice plants in upland rice fields at the Indian Agricultural Research Institute, New Delhi, India, were found severely stunted and chlorotic. Generally, these symptoms are due to iron deficiency. Further investigations revealed no iron deficiency. This prompted us to study the nematode infestation, if any, affecting aerobic rice plants. In these fields, the previous crop sequence was fallow followed by maize and then the aerobic rice cv. PRH 10 was direct seeded. Soil and root samples were collected 35 days after sowing.

A total of five soil samples (four from the corners and one from the centre) were collected from each of the three fields (A, B, C). Each sample was mixed in a 24-inch plastic bowl and a sub-sample of 200 cm³ was processed by Cobb's sieving and decanting method (Cobb, 1918) combined with modified Baerman funnel technique (Southey, 1986). Three 1-ml aliquots of the nematode water suspension were observed under a stereo-microscope and the nematodes counted. The predominant plant nematodes were of the genus *Pratylenchus*. They were killed by adding to the water suspension an equal amount of boiling water and fixed in 2.5% formalin. The nematode specimens were then hand-picked, processed by Seinhorst's method (1959) and mounted in dehydrated glycerine.

Infected yellow and stunted plants were carefully uprooted and the roots were washed gently in tap water and stained in a boiling 0.1% acid fuchsin solution. The stained roots (0.5 g) were observed under a stereo-microscope and the nematodes teased out with the help of needles and counted.

The nematodes were then observed (12 specimens) under a microscope and identified to species level according to the key of Singh and Gill (1986). Because of the presence of three lip annules, stylet length of 15-16.8 μ m, vulva position at 74-80 %, total body length of 0.46-0.69 mm, post-uterine sac of 11-30 μ m, spermatheca empty, lack of males, and truncate non-annulated tail, the specimens were identified as *P. thornei* Sher *et* Allen, 1953.

Symptoms of lesion nematode (as with most nematode-induced diseases) often go unrecognized initially as the above ground symptoms are often general symptoms of plant root stress. The nematode populations in soil and roots were 406 per 200 cm³ soil and 32 nematodes per 0.5 g roots in the severely damaged field C



Fig. 1. Damages caused by Pratylencus thornei to rice. Left, patchy growth of infected plants. Right, plants with chlorotic leaves.

(Fig. 1). Only 12 nematodes per 200 cm³ soil occurred in a nearby field B, previously left fallow, in which rice was showing normal growth and no chlorosis (Table I). Infested roots showed characteristic necrotic lesions (darkened areas of dead tissue) on the surface and throughout the cortex (Fig. 2). The lesions turned from reddish-brown to black and were spotty along the root surface. Root growth was reduced and small lesions also formed on young roots. The lesions may also lead to secondary infections by fungi or bacteria, thus increasing damage severity (Agrios, 2005). In turn, this results in poor plant growth, reduced yield or even complete crop failure.

Crop damage is related to the soil nematode population density at germination. Usually, the root damage, and possible eventual plant death, becomes obvious by 30-40 days after germination. Sowing of upland rice is done with the onset of first showers and seedling survival and yields are completely dependent on the progress of the rainy season. If plant death occurs in irrigated rice gap filling is possible, but this is not possible in upland rice in which plants having damaged roots will not recover because of the limited water resources.

Nombela *et al.* (1998) reported much lower populations (relative abundance 0.41 ± 0.06) of *P. thornei* fol-



Fig. 2. Specimens of *P. thornei* in rice roots showing dark lesions.

lowing summer fallow compared to wheat or vetch (relative abundance 0.87 ± 0.07). In our study, the rather high soil population of *P. thornei* in an aerobic rice field could be due to the preceding maize crop, compared to fallow land preceding aerobic rice in another two fields showing low nematode soil population densities.

Table I. Nematode population in three experimental fields.

Field	Previous crop	Nematode population	
		per 200 cc soil	per 0.5 g root
А	Cowpea	23 (4.89) ^b	3 (2.0) ^d
В	Fallow	12 (3.60) ^b	$00 (1.0)^{d}$
С	Maize	406 (20.17) ^a	32 (5.74) ^c

Figures in the same column followed by the same letter do not differ significantly at P = 0.05 according to Duncan's multiple range test.

Figures in parentheses are sqrt x+1 transformed values.

Pratylenchus thornei may easily spread in rice fields with water and soil and, therefore, there is high potential for the infested area in the country to expand, which would result in severe yield losses to rice, particularly under sub-optimal management conditions (Pankaj *et al.*, 2006a, b).

Because of the above, farmers should be advised to avoid crop sequences leading to large soil population densities of *P. thornei* before cultivating upland rice under tropical conditions. However, further investigations under Indian rice cropping conditions are suggested to obtain more insights on the ecology and management of the nematode.

ACKNOWLEDGEMENTS

The authors are grateful to Dr. Mangu Singh and Mr. Khajan Singh, Division of Nematology, IARI, New Delhi for their help in collecting and processing root and soil samples.

LITERATURE CITED

- Agrios G.N., 2005. *Plant pathology* (5th ed.), Academic Press, San Diego USA, 852 pp.
- Bouman B.A.M. and Tuong T.P., 2001. Field water management to save water and increase its productivity in irrigated lowland rice. *Agricultural Water Management, 49*: 11-30.
- Bouman B.A.M., Peng S., Castañeda A.R. and Visperas R.M., 2005. Yield and water use of irrigated tropical aerobic rice systems. *Agricultural Water Management*, 74: 87-105.
- Bouman B.A.M., Yang X.G., Wang H.Q., Wang Z.M., Zhao J.F. and Chen B., 2006. Performance of aerobic rice varieties under irrigated conditions in North China. *Field Crops Research*, 97: 53-65.
- Bridge J., Plowright R.A. and Peng D.L., 2005. Nematode parasites of rice. Pp. 87-130. *In*: Plant Parasitic Nematodes in Subtropical and Tropical Agriculture - Second edition (Luc M., Sikora R.A. and Bridge J., eds). CABI Publishing, Wallingford, UK.
- Cobb N.A., 1918. Estimating the nematode populations of soil. Circ. No. 1, USDA, Washington, USA, 48 pp.
- Di Vito M., Greco N. and Saxena M., 1992: Pathogenecity of *Pratylenchus thornei* on chickpea in Syria. *Nematologia Mediterranea*, 20: 71-73.
- Dwivedi K. and Upadhyay K.D., 1998. Nematode pests in rice-wheat-legume cropping systems in central Uttar Pradesh. Pp. 60-62. *In*: Nematode pests in rice-wheatlegume cropping systems: Proceedings of a Regional Training Course, 1-5 September 1997, CCS Haryana Agricultural University, Hisar, Haryana, India (Sharma S.B., Jo-

hansen C. and Midha S.K., eds). Rice-Wheat Consortium Paper Series 4. New Delhi, India: Rice-Wheat Consortium for the Indo-Gangetic Plains.

- MacLean J.L., Dawe D.C., Hardy B. and Hettel G.P., 2002. *Rice Almanac* (3rd ed.). IRRI, Los Baños, Laguna, Philippines, 253 pp.
- Nicol J.M. and Ortiz-Monasterio I., 2004. Effect of root lesion nematode on wheat yields and plant susceptibility in Mexico. *Nematology*, 6: 485-493.
- Nicol J.M., Davies K.A., Hancock T.W. and Fisher J.M., 1999. Yield loss caused by *Pratylenchus thornei* on wheat in South Australia. *Journal of Nematology*, 31: 367-376.
- Nombela G., Navas A. and Bello A., 1998. Effects of crop rotations of cereals with vetch and fallow on soil nematofauna in central Spain. *Nematologica*, 44: 63-80.
- Oteifa B.A., 1962. Species of root-lesion nematodes commonly associated with economic crops in the Delta of the U.A.R. *Plant Disease Reporter*, 46 : 572-575.
- Pankaj, Sharma H.K., Gaur H.S. and Singh A.K., 2006a. Effect of zero tillage on nematode fauna in rice-wheat cropping system. *Nematologia Mediterranea*, 34: 173-176.
- Pankaj, Ahlawat J.S. and Saha M., 2006b. Predominant nematode pests of rice nursery in North-Western India. Pp. 23-24. *In*: 2nd International Rice Congress, (9-13 Oct, 2006), ICAR, New Delhi, India.
- Prasad J.S., Panwar M.S. and Rao Y.S., 1987. Nematode problems of rice in India. *Tropical Pest Management, 33*: 127-136.
- Seinhorst J.W., 1959. A rapid method for the transfer of nematodes from fixative to anhydrous glycerine. *Nematologi ca*, 4: 67-69.
- Singh R.V. and Gill J.S., 1986. Species of the genus *Pratylenchus* Filipjev, 1936 with a key for identification. Technical Bulletin, AICRP on nematode pests of crops and their control, IARI, New Delhi, India, 34 pp.
- Southey J.F., 1986. Laboratory methods for work with plant and soil nematodes. Her Majesty's Stationery Office, London, UK, 202 pp.
- Tuong T.P. and Bouman B.A.M., 2003. Rice production in water-scarce environments. Pp. 53-67. *In*: Water Productivity in Agriculture: Limits and Opportunities for Improvements (Kijne J.W., Barker R. and Molden D., eds). CABI Publishing, Wallingford, UK.
- Wang H.Q., Bouman B.A.M., Zhao D., Wang C.G. and Moya P.F., 2002. Aerobic rice in northern China: opportunities and challenges. Pp. 143-154. *In*: Water-Wise-Rice Production. Proceedings of the International Workshop on Water-Wise Rice Production (Bouman B.A.M., Hengsdijk H., Hardy B., Bindraban P.S., Tuong T.P. and Ladha J.K., eds). International Rice Research Institute, Los Baños, Laguna, Philippines.
- Yang X.G., Bouman B.A.M., Wang H.Q., Wang Z.M., Zhao J.F. and Chen B., 2005. Performance of temperate aerobic rice under different water regimes in North China. *Agricultural Water Management*, 74: 107-122.